

### **Conservation of gene function in behaviour**

Christopher J. Reaume and Marla B. Sokolowski

*Phil. Trans. R. Soc. B* 2011 **366**, 2100-2110 doi: 10.1098/rstb.2011.0028

References	This article cites 97 articles, 27 of which can be accessed free http://rstb.royalsocietypublishing.org/content/366/1574/2100.full.html#ref-list-1
	Article cited in: http://rstb.royalsocietypublishing.org/content/366/1574/2100.full.html#related-urls
Subject collections	Articles on similar topics can be found in the following collections
	neuroscience (548 articles) behaviour (2206 articles) evolution (2943 articles)
Email alerting service	Receive free email alerts when new articles cite this article - sign up in the box at the top right-hand corner of the article or click here

To subscribe to Phil. Trans. R. Soc. B go to: http://rstb.royalsocietypublishing.org/subscriptions



Review

### **Conservation of gene function in behaviour**

### Christopher J. Reaume and Marla B. Sokolowski\*

Department of Biology, University of Toronto, Mississauga, Ontario, Canada, L5L 1C6

Behaviour genetic research has shown that a given gene or gene pathway can influence categorically similar behaviours in different species. Questions about the conservation of gene function in behaviour are increasingly tractable. This is owing to the surge of DNA and 'omics data, bioinformatic tools, as well as advances in technologies for behavioural phenotyping. Here, we discuss how gene function, as a hierarchical biological phenomenon, can be used to examine behavioural homology across species. The question can be addressed independently using different levels of investigation including the DNA sequence, the gene's position in a genetic pathway, spatial-temporal tissue expression and neural circuitry. Selected examples from the literature are used to illustrate this point. We will also discuss how qualitative and quantitative comparisons of the behavioural phenotype, its function and the importance of environmental and social context should be used in cross-species comparisons. We conclude that (i) there are homologous behaviours, (ii) they are hard to define and (iii) neurogenetics and genomics investigations should help in this endeavour.

Keywords: gene function; behaviour; conservation; homology; genetics; genomics

#### **1. INTRODUCTION**

It is well known that gene sequences are conserved even between distantly related species. For example, genes in the nematode Caenorhabditis elegans or the fly Drosophila melanogaster exhibit sequence similarity to versions of human genes. But does such DNA sequence similarity reflect a functionally conserved role for the genes in question? The answer is yes for developmental genes such as hox genes that specify anterior-posterior morphology in organisms from flies to mammals suggesting that hox genes had this function in a common ancestor of arthropods and chordates [1]. Here, we ask if evolutionary developmental (evo-devo) approaches can be extended to behavioural phenotypes that exhibit extensive plasticity and are subject to real-time interaction with the environment.

# 2. HOMOLOGY AND THE CONSERVATION OF BEHAVIOURAL TRAITS

A phenotype is homologous when two (or more) species share a common ancestor that exhibits the phenotype. Distinguishing between evolutionary conservation and convergence is challenging [2,3]. Descriptions of homology have traditionally been the purview of embryologists, anatomists and systematists. More recently, evo-devo biologists have addressed morphological homology and the role that gene function plays in specifying conserved phenotypes across species. Molecular phylogenies have been used as important baseline data for tests of homology and

\* Author for correspondence (marla.sokolowski@utoronto.ca).

morphology [4]. Identifying homologous behavioural phenotypes is challenging because (i) behaviour exhibits plasticity in response to the environment and (ii) behaviour can show homology at one level of biological organization (e.g. gene pathway), but not at another (e.g. neural circuitry; see below) [5].

Homologous behaviours are hard to define. In developmental biology, researchers consider the modular nature of an organisms' body plan (e.g. a limb or an organ) in order to relate morphological features to phenomena at the cellular level to patterns of gene expression [6]. Similar to this is the concept of endophenotypes, where complex behaviours are constructed from simpler components or modules [7-9]. Endophenotypes and their behavioural components are useful for approaching investigations of homology in behaviour. For example, 'courtship behaviour' in D. melanogaster comprises component behaviours, including orientation, tapping, wing extension and song [10]. Examining behavioural components is not only important for tractability and interspecific comparisons of behaviours but can also help identify species-specific modifications in complex behavioural phenotypes that might not have been apparent.

How can behavioural phenotypes be compared in species with drastically different morphology and natural history? We can start with similar categories of behaviour that are shared across species such as feeding, mating, parental care, aggression, learning, memory, circadian rhythms and sleep. Each category can be quantified using descriptions of the behaviours performed (e.g. for aggression: kick, lunge, punch, bite) as well as information about the frequency, duration and sequencing of behaviours. A significant challenge is to develop and standardize informative and unbiased assays of behaviour across species using a comparative approach [11]. Part of this challenge is

One contribution of 10 to a Theme Issue 'Evolutionary developmental biology (evo-devo) and behaviour'.

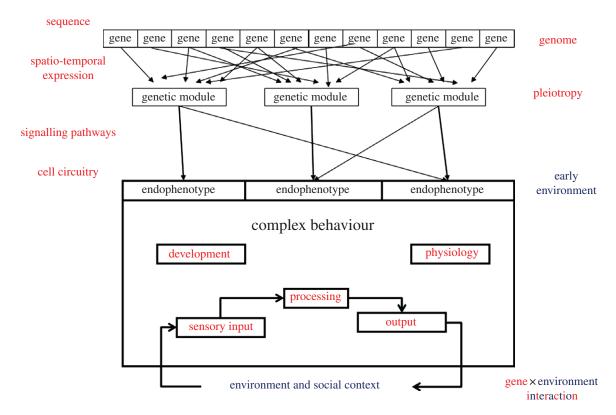


Figure 1. Schematic diagram of the hierarchical nature of gene function in behaviour. Included are a number of factors that can be examined for homology in influencing behaviour through gene function. Terms coloured in red indicate factors that are intrinsic to the organism that are involved in gene function in behaviour while those that are coloured blue are extrinsic to the organism. Epistasis and epigenetic processes are not shown in the figure; however, gene interaction networks and heritable changes in chromosome methylation and histone modification patterns that underlie behaviours also might be conserved across species.

to design paradigms that are relevant to the natural histories of each species but at the same time can be compared functionally across species [12]. This is more straightforward for some categories of behaviour than others (e.g. characteristics of rest/wake cycles compared with courtship behaviour). Considerations of the social context of behaviours should be incorporated into analyses as this information is inseparable from the phenotypes. Finally, the extent of plasticity of the behaviour is also important as it tells us whether and how much the phenotype varies across environments. These latter considerations introduce considerable analytical difficulties when comparing divergent species.

## 3. CONSERVATION OF GENE FUNCTION IN BEHAVIOUR

Information from neurogenetics and genomics helps determine which behaviours are homologous. Investigations of gene function homology in behaviour can be approached through interspecific comparisons of the various components that affect the behavioural phenotype in question [5,13,14]. The implicated genes, their sequence variation and the relevant signalling pathways and tissues (cells, organs, circuits) are all informative (see case studies below). In this sense, we define 'gene function in behaviour' as a hierarchical phenomenon that includes not only sequence identity and transcriptional events but also the position and role of the gene product in a signalling pathway that acts in defined cells and circuits in the expression of the behavioural phenotype (figure 1). Evo-devo studies have found that homologous morphological phenotypes may result from genetic and developmental mechanisms that are not themselves necessarily homologous [3,5]. This suggests that questions related to gene function homology should focus on a single hierarchical level of gene function (as it relates to behaviour) at a time. This is discussed further below.

We mentioned above the need for comparable and unbiased behavioural tests and well-known phylogenies for the species in question (see [12] for further discussion). Examples of other pertinent investigations include (i) genome or gene sequencing to test for sequence homology, (ii) transcriptional studies to assess gene expression, (iii) functional studies examining posttranslational and signalling pathways, (iv) comparative histological studies examining developmental and physiological aspects of the tissues and structures involved in gene function and execution of the behaviour in question, and (v) comparative studies examining gene-environment interactions of the gene(s) and behaviour(s) under investigation. This list is not exhaustive but provides several perspectives from which a gene's function in behaviour may be investigated. We suggest that the question of homology needs to be addressed and considered at each relevant level of gene function (cell, circuit, neural substrate, transcription, translation, signalling pathway, behaviour) independently (see case studies below).

Conservation of a gene's function can occur at the levels of molecular pathways, plasticity or gene– environment interactions, neural circuitry, developmental functions and through its pleiotropy (figure 1). What is meant here by conservation of a gene's pleiotropic function is when a gene affects the same suite of behaviours in two different species, suggesting shared pleiotropic functions of the gene in these species. Some scientists interested in genes and behaviour use the candidate gene approach to facilitate the identification of genes involved in the behaviours of a variety of species [15]. In this respect, candidate genes are those that are defined in one organism (often in well-defined genetic models like D. melanogaster and Mus musculus) and then investigated for similar effects in organisms without a genetic tool-box. Mutations in the genes of the former group of organisms are available or can be generated along with transgenic animals that can be used to increase or decrease expression of a gene and to target the expression of that gene in time and space. Numerous behaviours are studied in this way including courtship and mating, circadian rhythms, sleep, learning and memory, aggression, maternal behaviour and food-related behaviours [16,17]. The importance of olfaction, audition, taste, touch and other stimuli to these behaviours is also under investigation. In some cases, the neural substrates important to these behaviours have been identified and manipulated. Many genes that influence these behaviours have been discovered using analyses of genetic mutants. Natural genetic variants in behaviours have also been studied and genes that affect normal individual differences in behaviour have been uncovered (e.g. foraging in Drosophila [18]; npr1 in C. elegans [19,20]; vasopressin receptor in mice and voles [21]).

An important challenge to identifying homology of gene function in behaviour is that well-resolved phylogenies are lacking for many species, making it difficult to test alternative hypotheses [12]. Below, we examine several well-known and extensively studied examples of gene function that influence behaviour across species. It is difficult to decipher what definition of 'homology' remains once a system-based approach to gene function in behaviour is adopted. The case studies presented below show that there is no singular approach to assess homology, because examinations at different levels of the system can lead to different conclusions. This is not an exhaustive review of all pertinent examples but a selection of useful studies for illustrative purposes.

### 4. CASE STUDIES IN THE CONSERVATION OF GENE FUNCTION IN BEHAVIOUR

### (a) Gene-by-environment interactions and the serotonin transporter gene

Behaviour as a phenotype is highly responsive to the environment. This plasticity makes it particularly challenging for studies of homology. Despite the plasticity that emerges from the abiotic and biotic factors experienced by organisms during development and adulthood, it is still possible to find common patterns in their responses to environments. One way to investigate this is to use genetic variation to ask whether different genotypes differ in their sensitivity to the environment. One of the best examples of gene-by-environment interactions that apply across species involves allelic variation in the serotonin transporter gene (*5-HTT*) and its interaction with early experience [22]. *5-HTT* encodes for a protein involved in serotonin re-uptake. Studies of this gene in non-human primates and in human populations have identified a promoter-linked polymorphic region that interacts with early experience to affect behaviours in the young and adults [23]. The long and short alleles result from a 43 bp insertion/deletion in the promoter region of the 5-HTT gene. In humans, the short allele has approximately three times less in vitro basal transcription of 5-HTT mRNA when compared with the long allele [24]. The allelic variants are differently associated with depression and other related behaviours when an individual has a history of adversity early in life [22]. In general, the short allele is thought to confer risk to early adversity while the long allele confers protection (but this is not always the case [25]). Although rats do not have the long-short polymorphism in their 5-HTT promoter region, polymorphisms in the rat 5-HTT gene have also been found to interact with early experience to affect similar behaviours to those reported in rhesus monkeys and humans [26].

#### (b) Gene pathways and the biological clock

Genetic analyses of the functions of biological clocks help us to understand the conservation of molecular pathways in behaviour. The molecular pathways involved in circadian phenotypes, such as sleep/wake cycles, have been well-described in diverse organisms. Additionally, as mentioned previously, the behavioural outputs of the clock (e.g. activity, sleep/wake cycles) are relatively easily compared between divergent species (figure 2). First, we provide some background on the biological clock in *Drosophila*, where the genetic underpinnings of the clock were first discovered [28,29].

The period (per) gene, which affects circadian rhythm in D. melanogaster, has been used as a candidate gene to examine per homologues involved in other insects [28] and mammals [30]; per was the first gene discovered to affect circadian behaviour [31]. Three mutations called long (*per<sup>l</sup>*), short (*per<sup>s</sup>*) and arrhythmic (*per<sup>l</sup>*) alter eclosion rhythms and circadian patterns of locomotor activity. A second clock gene called timeless (tim) affects circadian rhythmicity and per expression [32-34]. Genes per and tim are transcriptionally regulated in a cyclic manner. Transcripts of both genes are present early in the day but the highest levels are found late in the day and at the beginning of the night [35–37]. PER and TIM proteins accumulate during the night and form a heterodimer that moves into the nucleus to bind to transcription factors Clock (CLK) and Cycle (CYC). This prevents CLK and CYC binding to the *per* and *tim* promoters which results in the transcriptional repression of *per* and *tim*. In early morning hours, TIM and PER degrade and allow for the rise in tim and per transcripts. This negative effect of PER and TIM on their own transcription creates the negative feedback loop that has been the central theme of clocks in many species [38]. Natural variants in per and tim are also known [39] and provide fertile ground for exploring gene function homology in behaviour. In fact, detailed comparisons among a number of different insect orders have already commenced [40]. There is general agreement that the function of *per* in biological rhythms is conserved across a broad range of species

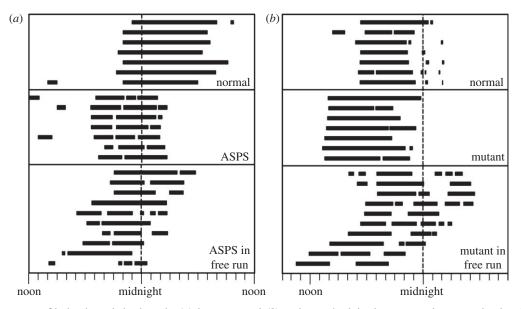


Figure 2. Actograms of behavioural rhythms in (a) humans and (b) rodents. Activity is measured as an episode of sleep (dark line) in the human and wheel running (dark line) in the rodent. Episodes of behaviour are shown relative to midnight with dark lines. Consecutive days are plotted top to bottom for each organism. The upper panels show behaviour rhythms of individuals maintained on light : dark cycles. The middle panels show a human with a sleep disorder called advanced sleep phase syndrome (ASPS) and a hamster with a mutation in the double-time gene which affects the biological clock. In the bottom panel the individuals are kept in constant darkness without any exposure to temporal cues. In both cases, the human with the sleep disorder and the mouse with a mutation in the period gene *Per2* show drift (from the normal 24 h period) in their rhythms. This drift is indicative of shortened periods. This figure illustrates the similarities in the circadian phenotypes of humans and rodents (figure adapted from [27]).

from insects to humans; however, *tim's* role in the biological clock is not. Gene and genome duplication events have produced four paralogues of *per* genes in mammals known as *mPer1* to *mPer4* [41]. The *mPer1* and *mPer2* genes appear to have a functionally similar role in the signalling pathways of flies and mammals. Circadian genes are thought to function in a number of human disorders [42].

Figure 3 shows the molecular pathways involved in clock function in the fly and the mouse. What appears to be conserved is the structure of the clock molecular mechanism but not necessarily each particular gene or gene product. (See also a discussion of relationship of the cvanobacterial clock to eukarvote clocks [43]). While some genes play the same roles in insects and mammals (e.g. period), others do not (timeless), and still others are found in some but not all species. Nevertheless, similar to well-known examples in evodevo, we can conclude that the structure of the molecular mechanism underlying clocks is similar in both groups. The genes involved are highly conserved at the DNA level, and some genes function in the same way and position in the clock molecular mechanism. Qualitative comparisons of figure 3 suggest that the raw material from the fly clock may have been 'tinkered with', in an evolutionary sense, to 'build' the mammalian clocks. If we were to ask whether there is conservation of a specific gene's function in behaviour, we would conclude that this is true for some genes and not others. For example, one could ask when and why the function of the *timeless* gene has changed over the course of vertebrate evolution. Or how genomic evolution has allowed for the potential conservation of the structure of clock molecular mechanisms while the body plan and organs in which it acts

have changed dramatically. In general, it is valuable to focus on divergences and convergences found between species at any level of organization as these cases will be informative from an evolutionary perspective and may suggest novel hypotheses.

To summarize this example, circadian rhythms are found in organisms from bacteria to humans, and these seem to be controlled by a 'clock' mechanism. Interestingly, while there is broad overlap between the 'genes' involved, they are not always the same. The evo-devo approach for circadian behaviour works well, and candidate gene approaches allowed us to make significant headway in understanding the genetic basis of circadian behaviour. However, the precise details of clock function require the study of clock mechanisms in specific organisms.

(c) Regulation of the foraging gene across species Even when sequence homology is found, as is required for the candidate gene approach, the details of gene function may differ at any or all hierarchical levels. The candidate gene approach [14] has been successfully used for studies of the D. melanogaster foraging (for) gene, investigated in C. elegans, Apis mellifera and the ants Pogonomyrmex barbatus and Pheidole pallidula (see below). The for gene encodes a cGMPdependent protein kinase (PKG) in D. melanogaster and affects a large array of behaviours including food-related behaviours, responses to stress and learning and memory [44]. Foraging behaviour in insects has been a major focus of research on the behavioural effects of PKG. In nature, larval and adult flies behave as rovers or sitters [45]. Well-fed rovers exhibit significantly more locomotion in the presence of food than



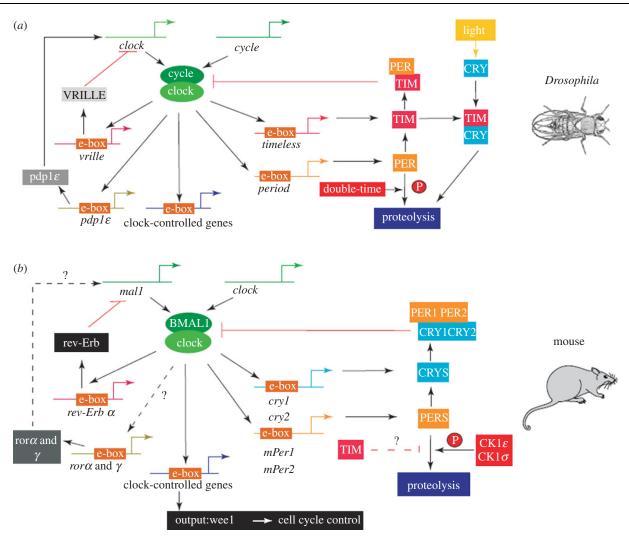


Figure 3. The molecular mechanism of the (a) Drosophila and (b) mammalian clocks. The genes that show sequence homology are shown in the same colour, those that are not homologous are shown in shades of grey. Proteins that form dimerization partners are presented as a single complex (figure adapted from [41]).

sitters. When food is patchy, rovers have a greater tendency to leave a patch in search of new food patches compared with sitters [15]. Food deprived rover larvae behave as sitters and exhibit concomitant reductions in their PKG enzyme activity resembling well-fed sitters [46,47]. In addition, well-fed rover larvae have lower food intake than sitters. Adult rovers and sitters show flexible responses to food deprivation as reflected in behavioural, metabolomic and microarray data [48,49]. As mentioned above, the for gene also plays a role in food-related behaviours in C. elegans, D. melanogaster, A. mellifera, P. barbatus and P. pallidula [50-53]. However, the particular functions of for and the signalling pathways involved must differ between species in significant ways. In D. melanogaster and A. mellifera, high levels of for result in rover and forager flies and bees, respectively. In C. elegans and P. pallidula, high levels of for result in dweller (not roamer) worms, and forager (not defender) ants, respectively. Furthermore, for's contribution to the plasticity of behaviour differs between these species. In D. melanogaster, both chronic food deprivation and short-term acute deprivation changes for expression and behaviour [46,47]. In A. mellifera and P. barbatus, for is involved in long-term plastic

changes in behaviour tied to maturation (temporal polyphenism). In P. pallidula, for is involved in a worker ant's rapid switch from defending the nest to foraging. Finally, only in *P. pallidula* has a difference in the spatial localization of FOR protein been reported. The brains of defender worker ants have five more cells expressing FOR than do the brains of the forager worker ants [53]. Together, these differences suggest that in a very broad sense, for is involved in the plasticity of food-related behaviours, but the actual time scale and mechanisms involved in the various species probably differ. It seems clear, however, that for modulates many phenotypes. It is not known whether for affects the same suite of phenotypes in other species and, if so, whether the multiple functions of a given gene (i.e. its pleiotropy) could be useful for between species comparisons. Further research should shed light on these issues.

The *for* gene example shows that a given gene or set of genes can contribute to phenotypic plasticity in behaviour across a diverse range of species [54]. This suggests, as in the biological clock example above, that the gene–gene interactions may also show conservation. The gene *for* and genes in the insulin signalling pathways are involved in both rover/sitter foraging [48] and the transition from nurse to forager in *A. mellifera* where the behavioural phenotypes and patterns of gene expression are nutritionally regulated.

#### (d) Genetic analyses of learning and memory

Another field of research, learning and memory, helps shed light on the conservation of gene function in behaviours across divergent species. The many types of learning and memory will not be discussed here nor will their relevance for the species' life histories. There are many paradigms for testing learning and memory, both non-associative and associative, that are informative for interspecific comparisons. One non-associative paradigm is habituation distinguished by decreases in a response to a stimulus following repeated stimulation. Another is sensitization, an increase in response to a neutral stimulus following exposure to a stronger stimulus. Habituation and sensitization are observed in species as diverse as C. elegans [55], D. melanogaster [56], Rattus norvegicus [57] and humans [58]. Associative learning has also been observed in many species [59,60].

Associative learning paradigms can be separated into two categories: (i) operant and (ii) classical (Pavlovian) conditioning [60]. In operant conditioning, an animal acts on its environment to establish associations between unrelated stimuli. The concept can easily be demonstrated by imagining an operant conditioning chamber ('Skinner box'). In one of its most basic forms, the operant conditioning chamber contains a lever that an animal learns to press in order to acquire a food reward. Classical conditioning usually involves the pairing of two stimuli. There is a reflexive stimulus that elicits a response (e.g. food reward, electric shock) and a 'conditioned stimulus' (e.g. an odour). The animal learns to associate the conditioned stimulus with the reflexive stimulus resulting in a conditioned response. These and other forms of non-associative and associative learning can be used to address questions of homology of gene function. However, some forms will be more tractable than others. For example, in operant conditioning, there is less control over the timing, duration and strength of the stimuli when compared with classical conditioning. Another consideration is the divergence of the organisms in question. Simpler learning paradigms such as habituation and sensitization are more suitable for addressing questions about the conservation of gene function.

Similarities in the behavioural properties of classical conditioning are found in a wide array of species from molluscs to insects, fish, rodents and humans. Many of the genetic pathways underlying memory formation are shared in these species [61,62]. For example, the cAMP-dependent signalling pathway has been implicated in memory formation in *Aplysia* [63], honey bees [64], flies [65] and rodents [66] as well as other species. More specifically, CREB (cAMP responsive element binding protein) is thought to be necessary for the formation of protein synthesis-dependent long-term memory in flies and rodents [67–70]. The importance of the cAMP signalling pathway for learning and memory is well established [71]. Some recent

Phil. Trans. R. Soc. B (2011)

investigations in *Drosophila* and mice show that PKG (called cGKI in mammals) also functions in learning and memory. In the mouse, PKG plays a role in fear conditioning in the amygdala [72]. In *Drosophila*, PKG is encoded by *for* with the rover and sitter natural variants described above. *for* is the fly homologue of mammalian cGK1 and is known to affect Pavlovian associative olfactory aversive learning [73,74], appetitive learning [46] and operant visual learning [75].

Many of the genes and signalling pathways associated with memory formation are conserved across a wide range of species. Drosophila genes involved in cAMP signalling, ras/MAP kinase signalling, Staufen RNA binding protein, and genes involved in human neurocognitive disorders all play a role in memory formation [61,62]. Furthermore, structural elements of the neural circuitry underlying associative learning are postulated to be homologous in insects and mammals [61]. Imaging analyses have advanced to the point where a common origin between the annelid mushroom bodies and the vertebrate pallium has been suggested [76]. Technological advancements in functional neuroanatomy and real-time imaging improve along with our understanding of the cellular mechanisms underlying learning and memory, and will facilitate further interspecific studies of learning and memory [71].

#### (e) Sleep

Sleep is another good model to examine homology of gene function in behaviour [77–80]. A number of behavioural criteria can be used to compare sleep between species [81]. They were designed to distinguish sleep from other states of quiescence and include: (i) a quiescent period, (ii) a reduction in the response to external stimuli (increased arousal threshold), (iii) increased rest after prolonged waking, and (iv) reversibility. These criteria led to the discovery of sleep states in a number of non-mammalian species including *C. elegans* [82], *Leucophea maderae* [83], *A. mellifera* [84], *D. melanogster* [85,86] and *Danio rerio* [87].

A cAMP-CREB pathway has been implicated in sleep, providing examples of various components of hierarchical gene function homology in flies, worms and mammals [78-80]. In Drosophila, mutants with increased cAMP levels have reduced sleep, while mutants with reduced cAMP levels have increased sleep [88]. Additionally, manipulations of CREB activity demonstrated its role in wakefulness. Further studies showed mice lacking either one or two CREB isoforms exhibited reduced wakefulness [89]. In worms, the mutants pde-4 (reduced-function cyclic nucleotide phosphodiesterase) and acy-1 (gain-of-function adenelyte cyclase) result in increased cAMP levels, and show increase sensory responsiveness during lethargus [82]. These data suggest that the role of cAMP signalling in sleep behaviour may be homologous at multiple levels in very diverse organisms.

Interestingly, the cGMP pathway, involving PKG, has been implicated in sleep-like behaviour. Raizen *et al.* [82] showed that PKG regulates sleep-like behaviours in flies and worms (*for* and *egl-4*, respectively). The authors compared gain- and loss-of-function

egl-4 mutants and demonstrated that PKG is associated with the extent of behavioural quiescence as well as its time-dependence. They then used rover and sitter fly lines to ask whether the behavioural effect of PKG on sleep is evolutionarily conserved. They found that rovers with higher PKG activity slept more than the sitters. These preliminary data suggest that PKG activity is positively associated with the amount of sleep that an animal displays in both species and appears to be conserved. In mice, a conditional knockout of mammalian PKG called cGKI results in increased sleep fragmentation, exaggerated delta rebound following deprivation and reduced rapid-eye movement sleep [90]. The cAMP and cGMP signalling pathways are important to sleep in diverse animals.

#### (f) Dopamine and reward

A given gene, or set of genes, may play a role in development and/or functioning of the neural circuitry of a behaviour phenotype. If this neural circuitry shows some conservation between species, then this circuitry can be investigated for conservation of behavioural function. Dopamine signalling regulates a variety of complex behaviours in a wide range of organisms [91]. The dopamine system is of interest because it functions in reward which is intimately linked with many behaviours such as feeding, mothering, sex, learning and addictive behaviours. Dopamine neurons express dopamine pathway genes whose products are involved in dopamine synthesis and transport in most organisms. All dopamine neurons share a small number of genes that code for enzymes and transporters important for the synthesis, packaging and re-uptake of dopamine. How these genes are regulated in diverse species is poorly understood. Flames & Hobert [91] recently found that the function of a dopamine *cis* regulatory motif called DA is conserved (and interchangeable) in C. elegans and M. musculus. These and other findings will open the door towards understanding the evolution of structures and neural circuits in animal brains [92].

#### (g) Neuropeptides and social behaviour

As discussed, questions about the conservation of behavioural phenotypes across distantly related species are difficult to answer. Donaldson & Young [93] discuss how vasopressin and its receptors play a role in the modulation of social and reproductive behaviours, a broad class of behaviours found in many organisms. However, the actual effects of this neuropeptide on components of these behaviours are highly speciesspecific. In closely related vole species, species-specific differences in the social bonding result from differences in the expression of the arginine vasopressin V1a receptor (V1aR). Monogamous prairie vole males have a higher number of V1a receptors than polygamous meadow vole males [21]. These voles differ in sequence variation at the 5' region of this gene. Genetic polymorphisms in this gene have also been associated with variation in sociobehavioral traits in humans, including autism spectrum disorders. However, the evolution of the 5' region of this vasopressin receptor gene did not directly contribute to

variation in social behaviour investigated in 13 species of primates [94]. It is difficult to determine whether the similarities in the behavioural functions of the vasopressin molecule and its receptors across distantly related species are due to homology. A category of behaviour that includes social and reproductive behaviours may be too broad to be used to address questions of homology. This suggests that the level of behavioural analysis (e.g. endophenotypes, behavioural components) that is chosen for investigations of homology can affect the conclusions.

When vasopressin is investigated as a class of molecules, a great deal of conservation of the neural expression of these genes is found [93]. DNA sequence homologues of this neuropeptide found in animals from hydra to vertebrates existed hundreds of millions of years ago. Examination of tissue-specific expression patterns shows that in mammals, vasopressin is found in the hypothalamic brain regions and then in the pituitary where it travels to affect the brain and is also released into the periphery. Interestingly, genetic homologues of vasopressin are found in related neurosecretory structures in the brains of other organisms such as worms and fish. So at the tissue level, this neuropeptide is found in functionally related regions and tissue types in mammals, fish and worms.

One way to ask whether the function of a gene has been conserved is to perform transgenic experiments between species. Transgenic experiments, using another neuropeptide called oxytocin, were performed to ask whether genes expressed in evolutionarily conserved neural tissue exhibit similar functions across species. Indeed transgenic rats carrying a blowfish oxytocin homologue were able to express oxytocin in neurons of the rat hypothalamus; this suggested consistency between the regulatory features of the blowfish and rat genes [95]. Additionally, the fish gene in transgenic rats exhibited normal physiological functions. While the behaviours investigated are highly species-specific, the levels of DNA sequence, tissue-specific expression, regulation and physiological function appear to be conserved in these neuropeptides. This example demonstrates that different levels of investigation can provide different insights into questions of gene function homology across diverse species.

## (h) Epigenetics, genomic responses to early adversity

A relatively new area of research that can potentially be scrutinized for gene function conservation in behaviour is epigenetics. For instance, specific environmental factors that alter methylation patterns or histone modification of conserved genes that affect behaviour might be homologous across diverse taxa. Meaney, Szyf and colleagues have characterized an epigenetic mechanism which results in 'maternal programming' that has been shown to affect stress responses in rats [96–98]. This response is mediated by glucocorticoid receptors in the hippocampus. Maternal licking and grooming provided by the mothers during the first week of life change the levels of RNA expression of the glucocorticoid receptors. High licking and grooming mothers produce offspring with higher levels of

glucocorticoid receptor mRNA, while low licking and grooming mothers produce offspring with lower levels of this RNA. Individuals that received higher maternal stimulation showed less behavioural and neuroendocrine reactivity to stressful stimuli. These changes in gene activity in response to stress are controlled by patterns of methylation that define an epigenetic response to mothering. The glucocorticoid receptor is affected by patterns of DNA methylation within the promotor region of the gene. In the case of low licking and grooming mothers, the offspring's promoter of the glucocorticoid receptor is methylated resulting in a decrease in the expression of this gene. This does not occur in offspring of high licking and grooming mothers. The patterns of methylation are maintained into later stages of life. There is some evidence that the stress response-related effects of maternal licking and grooming are passed on to female offspring. The female offspring mother their offspring according to how they themselves were reared (with high or low licking and grooming). The transmission of this maternal behaviour across generations is related to methylation of the oestrogen receptor gene passed from mother to daughter [98]. In a related human study, McGowan et al. [99] showed that methylation patterns and RNA expression levels of the glucocorticoid receptor in the hippocampus of the brains of suicide victims were altered when the victims had a history of abuse. This suggested that analogous to low licking and grooming in mice, early adversity in humans causes a downregulation of the glucocorticoid receptor gene. Thus, this gene-by-early environment adversity interaction is found in rats and humans.

#### 5. SUMMARY

Early on, the field of behaviour genetics focused on organisms that could be genetically manipulated. However, this limited the breadth of species and behaviours that could be studied using a genetic approach. Advances in DNA sequencing and the 'omics sciences as well as the use of candidate genes has allowed for a broader focus that includes new model species studied from ecological and neurobiological perspectives. Indeed, it has been argued that an understanding of how genes evolve to affect phenotypes should include a comparative approach, and should consider many species and collaborations between evolutionary and molecular biologists [100,101]. While relatively little is known about how evolutionary processes shape intra- and interspecific variation in behavioural phenotypes and the genes that underlie them [7,8], one intriguing theme is that genes which show homology at the level of DNA sequence appear to influence similar categories of behaviours across taxa.

What can functional genomics, systems biology and the plethora of data provided [102] tell us about conservation of a gene's function in behaviour? As the sophistication of genome databases rapidly improve, we will learn more about molecular pathways involved in specific brain functions and how these pathways translate to species-specific differences in behaviour. By making comparisons across genomes, we can better understand how sequence variation, genetic architecture and expression patterns associate with

conserved phenotypes. Evo-devo has already used genome-wide linkage mapping and transcriptional profiling for interspecific comparisons. Such genome-wide comparisons can be measured dynamically and in parallel between multiple species of interest. Combined with information on nervous system function, neural circuitry and plasticity, we will be able to compare and contrast molecular pathways and their neural substrates for well-defined behavioural phenotypes across species in different environments. This system-level approach will provide data to address issues of conservation of the molecular and neural pathways that underlie specific behavioural variation. Rapid developments should soon make it possible to link levels of organization on a genome and nervous system-wide scale, making it possible to address issues of conservation across all levels of organization in more quantitative ways.

We thank Dr James Burns, Dr Ken Dawson-Scully and Dr Scott Douglas for fruitful discussions realting to the topics discussed in the article, Dr Laurence Packer for useful comments on an early version of the manuscript and Bianco Marco for help with preparation of the manuscript. We also especially thank Rinaldo Bertossa and three anonymous reviewers for critical readings and very useful suggestions that improved the article. This work was supported by Natural Sciences and Engineering Research Council of Canada (NSERC) grants to Marla B. Sokolowski and Christopher J. Reaume.

#### REFERENCES

- 1 Caroll, S. B., Grenier, J. K. & Weatherbee, S. D. 2005 From DNA to design: molecular genetics and the evolution of animal design. Oxford, MA: Blackwell Science.
- 2 Zakon, H. H. 2002 Convergent evolution on the molecular level. *Brain Behav. Evol.* 59, 250–261. (doi:10. 1159/000063562)
- 3 Hall, B. K. 2003 Descent with modification: the unity underlying homology and homoplasy as seen through an analysis of development and evolution. *Biol. Rev.* 78, 409–433. (doi:10.1017/S1464793102006097)
- 4 Gibson, G. & Muse, S. V. 2009 Genome sequencing and annotation. In *A primer of genome science*, pp. 65–131. Oxford, MA: Sinauer Associates.
- 5 Hall, B. K. 2007 Homoplasy and homology: dichotomy or continuum? *J. Hum. Evol.* **52**, 473–479. (doi:10. 1016/j.jhevol.2006.11.010)
- 6 Wagner, G. P., Pavlicev, M. & Cheverud, J. M. 2007 The road to modularity. *Nat. Rev. Genet.* 8, 921–931. (doi:10.1038/nrg2267)
- 7 Barron, A. B. & Robinson, G. E. 2008 The utility of behavioral models and modules in molecular analyses of social behavior. *Genes Brain Behav.* 7, 257–265. (doi:10.1111/j.1601-183X.2007.00344.x)
- 8 Toth, A. L. & Robinson, G. E. 2007 Evo-devo and the evolution of social behavior. *Trends Genet.* 23, 334–341. (doi:10.1016/j.tig.2007.05.001)
- 9 Wenzel, J. W. 1993 Application of the biogenetic law to behavioural ontogeny: a test using nest architecture in paper wasps. *J. Evol. Biol.* 6, 229–247. (doi:10.1046/j. 1420-9101.1993.6020229.x)
- 10 Greenspan, R. J. & Ferveur, F. 2000 Courtship in Drosophila. Annu. Rev. Genet. 34, 2005–2232. (doi:10. 1146/annurev.genet.34.1.205)
- 11 Tinbergen, N. 1951 *The study of instinct*. New York, NY: Oxford University Press.
- 12 Pollen, A. A. & Hofmann, H. A. 2008 Beyond neuroanatomy: novel approaches to studying brain evolution.

2108 C. J. Reaume & M. B. Sokolowski Review. Gene function conservation in behaviour

Brain Behav. Evol. 72, 145–158. (doi:10.1159/000151474)

- 13 Rendall, D. & Di Fiore, A. 2007 Homoplasy, homology, and the perceived special status of behavior in evolution. *J. Hum. Evol.* 52, 504–521. (doi:10.1016/j.jhevol.2006. 11.014)
- 14 Wenzel, J. W. 1992 Behavioral homology and phylogeny. Annu. Rev. Eco. Syst. 23, 361–381. (doi:10.1146/ annurev.es.23.110192.002045)
- 15 Fitzpatrick, M. J., Ben-Shahar, Y., Smid, H. M., Vet, L. E. M., Robinson, G. E. & Sokolowski, M. B. 2005 Candidate genes for behavioural ecology. *Trends Ecol. Evol.* **20**, 96–104. (doi:10.1016/j.tree. 2004.11.017)
- 16 Sokolowski, M. B. 2001 Drosophila: genetics meets behaviour. Nat. Rev. Genet. 2, 879–892. (doi:10.1038/ 35098592)
- 17 Sokolowski, M. B. 2010 Social interactions in 'simple' model systems. *Neuron* 65, 780–794. (doi:10.1016/j. neuron.2010.03.007)
- 18 Osborne, K. A., Robichon, A., Burgess, E., Butland, S., Shaw, R. A., Coulthard, A., Pereira, H. S., Greenspan, R. J. & Sokolowski, M. B. 1997 Natural behavior polymorphism due to a cGMP-dependent protein kinase of *Drosophila. Science* 277, 834–836. (doi:10.1126/science. 277.5327.834)
- 19 de Bono, M. & Bargmann, C. I. 1998 Natural variation in a neuropeptide Y receptor homolog modified social behaviour and food response in *C. elegans. Cell* 4, 679–689. (doi:10.1016/S0092-8674(00)81609-8)
- 20 McGrath, P. T., Rockman, M. V., Zimmer, M., Jang, H., Macosko, E. Z., Kruglyak, L. & Bargmann, C. I. 2009 Quantitative mapping of a digenic behavioral trait implicates globin variation in *C. elegans* sensory behaviors. *Neuron* **12**, 692–699. (doi:10.1016/j.neuron. 2009.02.012)
- 21 Young, L. J., Wang, Z. & Insel, T. R. 1998 Neuroendocrine bases of monogamy. *Trends Neurosci.* 21, 71–75. (doi:10.1016/S0166-2236(97)01167-3)
- 22 Caspi, A. *et al.* 2003 Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science* **301**, 386–389. (doi:10.1126/science.1083968)
- 23 Murphy, D. L., Fox, M. A., Timpano, K. R., Moya, P. R., Ren-Patterson, R., Andrews, A. M., Holmes, A., Lesch, K. P. & Wendland, J. R. 2008 How the serotonin story is being rewritten by new gene-based discoveries principally related to SLC6A4, the serotonin transporter gene, which functions to influence all cellular serotonin systems. *Neuropharma* 55, 932–960. (doi:10.1016/j.neuropharm.2008.08.034)
- 24 Heils, A., Teufel, A., Petri, S., Stöber, G., Riederer, P., Bengel, D. & Lesch, K. P. 1996 Allelic variation of human serotonin transporter gene expression. *J. Neurochem.* 66, 2621–2624. (doi:10.1046/j.1471-4159.1996.66062621.x)
- 25 Mileva-Seitz, V., Kennedy, J., Atkinson, L., Steiner, M., Levitan, R., Matthews, S. G., Meaney, M. J., Sokolowski, M. B. & Fleming, A. S. 2011 Serotonin transporter allelic variation in mothers predicts maternal sensitivity, behaviour and attitudes toward 6-month-old infants. *Genes Brain Behav.* 10, 325–333. (doi:10.1111/j.1601-183X.2010.00671.x)
- 26 Belay, H., Burton, C. L., Vedran, L., Meaney, M. J., Sokolowski, M. B. & Fleming, A. S. 2011 Early adversity and serotonin transporter genotype interact with hippocampal glucocorticoid receptor mRNA expression, corticosterone, and behavior in adult male rats. *Behav. Neurosci.* **125**, 150–160. (doi:10.1037/ a0022891)

- 27 Wager-Smith, K. & Kay, S. A. 2000 Circadian rhythm genetics: from flies to mice to humans. *Nat. Genet.* 26, 23–27. (doi:10.1038/79134)
- 28 Hall, J. C. 2003 Genetics and molecular biology of rhythms in *Drosophila* and other insects. *Adv. Genet.* 48, 1–280. (doi:10.1016/S0065-2660(03)48000-0)
- 29 Boothroyd, C. E. & Young, M. W. 2008 The in(put)s and out(put)s of the *Drosophila* circadian clock. *Ann. NYAcad. Sci.* **1129**, 350–357. (doi:10.1196/annals.1417.006)
- 30 Panda, S., Hogenesch, J. B. & Kay, S. A. 2002 Circadian rhythms from flies to humans. *Nature* 417, 329–335. (doi:10.1038/417329a)
- 31 Konopka, R. J. & Benzer, S. 1971 Clock mutants of Drosophila melanogaster. Proc. Natl Acad. Sci. USA 68, 2112–2116. (doi:10.1073/pnas.68.9.2112)
- 32 Sehgal, A., Price, J. L., Man, B. & Young, M. W. 1994 Loss of circadian behavioral rhythms and *per* RNA oscillations in the *Drosophila* mutant timeless. *Science* 263, 1063–1606. (doi:10.1126/science.8128246)
- 33 Voshall, L. B., Price, J. L., Sehgal, A., Saez, L. & Young, M. W. 1994 Block in nuclear localization of *period* protein by a second clock mutation, *timeless. Science* 263, 1606-1609. (doi:10.1126/science.8128247)
- 34 Myers, M. P., Wager-Smith, K., Rothenfluh-Hilfiker, A. & Young, M. W. 1996 Light-induced degradation of TIMELESS and entrainment of the *Drosophila* circadian clock. *Science* 271, 1736–1740. (doi:10.1126/science. 271.5256.1736)
- 35 So, W. V. & Rosbash, M. 1997 Post-transcriptional regulation contributes to *Drosophila* clock gene mRNA cycling. *EMBO J.* 16, 146–155. (doi:10.1093/emboj/ 16.23.7146)
- 36 Hardin, P. E., Hall, J. C. & Rosbash, M. 1990 Feedback of the *Drosophila period* gene on circadian cycling of its messenger RNA levels. *Nature* 343, 536–540. (doi:10. 1038/343536a0)
- 37 Sehgal, A., Rothenfluh-Hilfiker, A., Hunter-Ensor, M., Chen, Y., Myers, M. P. & Young, M. W. 1995 Rhythmic expression of *timeless*: a basis for promoting circadian cycles in *period* gene autoregulation. *Science* 270, 808-810. (doi:10.1126/science.270.5237.808)
- 38 Rosato, E. & Kyriacou, C. P. 2001 Flies, clocks and evolution. *Phil. Trans. R. Soc. Lond. B* 356, 1769–1778. (doi:10.1098/rstb.2001.0961)
- 39 Kyriacou, C. P., Peixoto, A. A., Sandrelli, F., Costa, R. & Tauber, E. 2007 Clines in clock genes: fine-tuning circadian rhythms to the environment. *Trends Genet.* 24, 124–132. (doi:10.1016/j.tig.2007.12.003)
- 40 Sandrelli, F., Costa, R., Kyriacou, C. P. & Rosato, E. 2008 Comparative analysis of circadian clock genes in insects. *Insect Mol. Biol.* **17**, 447–463. (doi:10.1111/j. 1365-2583.2008.00832.x)
- 41 Looby, P. & Loudon, A. S. I. 2005 Gene duplication and complex circadian clocks in mammals. *Trends Genet.* 21, 46–53. (doi:10.1016/j.tig.2004.11.012)
- 42 Barnard, A. & Nolan, P. M. 2008 When clocks go bad: neurobehavioural consequences of disrupted circadian timing. *PLoS Genet.* 4, e1000040. (doi:10.1371/ journal.pgen.1000040)
- 43 Rosbash, M. 2009 The implications of multiple circadian clocks origin. *PLoS Biol.* 7, e1000062. (doi:10. 1371/journal.pbio.1000062)
- 44 Reaume, C. J. & Sokolowski, M. B. 2009 cGMPdependent protein kinase as a modifier of behaviour. *cGMP: generators, effects and therapeutic implications. H.E.P.* 191, 423–443. (doi:10.1007/978-3-540-68964-5\_18)
- 45 Sokolowski, M. B., Pereira, H. S. & Hughes, K. 1997 Evolution of foraging behavior in *Drosophila* by density-dependent selection. *Proc. Natl Acad. Sci. USA* 94, 7373–7377. (doi:10.1073/pnas.94.14.7373)

- 46 Kaun, K. R., Hendel, T., Gerber, B. & Sokolowski, M. B. 2007 Natural variation in *Drosophila* larval reward learning and memory due to a cGMP-dependent protein kinase. *Learn. Mem.* 14, 342–349. (doi:10.1101/lm.505807)
- 47 Kaun, K. R., Chakaborty-Chatterjee, M. & Sokolowski, M. B. 2008 Natural variation in plasticity of glucose homeostasis and food intake. *J. Exp. Biol.* 211, 3160– 3166. (doi:10.1242/jeb.010124)
- 48 Kent, C. F., Daskalchuk, T., Cook, L., Sokolowski, M. B. & Greenspan, R. J. 2009 The *Drosophila foraging* gene mediates adult plasticity and gene–environment interactions in behaviour, metabolites, and gene expression in response to food deprivation. *PLoS Genet.* 5, e1000609. (doi:10. 1371/journal.pgen.1000609)
- 49 Belay, A. T., Scheiner, R., So, A. K., Douglas, S. J., Chakaborty-Chatterjee, M., Levine, J. D. & Sokolowski, M. B. 2007 The *foraging* gene of *Drosophila melanogaster*: spatial-expression analysis and sucrose responsiveness. *J. Comp. Neurol.* **504**, 570–582. (doi:10.1002/cne.21466)
- 50 Fujiwara, M., Sengupta, P. & McIntire, S. L. 2002 Regulation of body size and behavioral state of *C. elegans* by sensory perception and the EGL-4 cGMP-dependent protein kinase. *Neuron* 36, 1091–1102. (doi:10.1016/S0896-6273(02)01093-0)
- 51 Ben-Shahar, Y., Robichon, A., Sokolowski, M. B. & Robinson, G. E. 2002 Influence of gene action across different time scales on behavior. *Science* 296, 741–744. (doi:10.1126/science.1069911)
- 52 Ingram, K. K., Oenfner, P. & Gordon, D. M. 2005 Task-specific expression of the foraging gene in harvester ants. *Mol. Ecol.* 14, 813–818. (doi:10.1111/j. 1365-294X.2005.02450.x)
- 53 Lucas, C. & Sokolowski, M. B. 2009 Molecular basis for changes in behavioral state in ant social behaviors. *Proc. Natl Acad. Sci. USA* **106**, 6351–6356. (doi:10. 1073/pnas.0809463106)
- 54 Ament, S. A., Corona, M., Pollock, H. S. & Robinson, G. E. 2008 Insulin signaling is involved in the regulation of worker division of labor in honey bee colonies. *Proc. Natl Acad. Sci. USA* **105**, 4226–4231. (doi:10.1073/ pnas.0800630105)
- 55 Rose, J. K. & Rankin, C. H. 2001 Analyses of habituation in *Caenorhabditis elegans*. Learn. Mem. 8, 63–69. (doi:10.1101/lm.37801)
- 56 Duerr, J. S. & Quinn, W. G. 1982 Three Drosophila mutations that block associative learning also affect habituation and sensitization. Proc. Natl Acad. Sci. USA 79, 3646–3650. (doi:10.1073/pnas.79.11. 3646)
- 57 Davis, M. 1970 Effects of interstimulus interval length and variability on startle-response habituation in the rat. *J. Comp. Physiol. Psychol.* 72, 177–192. (doi:10. 1037/h0029472)
- 58 Geer, J. H. 1966 Effect of interstimulus intervals and rest-period length upon habituation of the orienting response. *J. Exp. Psychol.* 72, 617–619. (doi:10.1037/ h0023760)
- 59 Shettleworth, S. J. 1998 Cognition, evolution, and behavior. New York, NY: Oxford University Press.
- 60 Bouton, M. E. 2007 Learning and adaptation. In Learning and behavior: a contemporary synthesis, pp. 39-72. Oxford, MA: Sinauer Associates.
- 61 Dubnau, J. 2003 Neurogenetic dissection of conditioned behavior: evolution by analogy or homology? *J. Neurogenet.* 17, 295–326. (doi:10.1080/01677060 390441859)
- 62 Davis, R. L. 2005 Olfactory memory formation in *Drosophila*: from molecular to systems neuroscience.

*Annu. Rev. Neurosci.* **28**, 275–302. (doi:10.1146/ annurev.neuro.28.061604.135651)

- 63 Kandel, E. R. 2001 The molecular biology of memory storage: a dialogue between genes and synapses. *Science* 294, 1030–1038. (doi:10.1126/science.1067020)
- 64 Eisenhardt, D. 2006 Learning and memory formation in the honeybee (*Apis mellifera*) and its dependency on the cAMP-protein kinase A pathway. *Anim. Biol.* 56, 259–278. (doi:10.1163/157075606777304249)
- 65 Margulies, C., Tully, T. & Dubnau, J. 2005 Deconstructing memory in Drosophila. Curr. Biol. 15, R700–R713. (doi:10.1016/j.cub.2005.08.024)
- 66 Josselyn, S. A., Shi, C., Carlezon Jr, W. A., Neve, R. L., Nestler, E. J. & Davis, M. 2001 Long-term memory is facilitated by cAMP response element-binding protein overexpression in the amygdala. *J. Neurosci.* 21, 2404–2412.
- 67 Yin, J. C., Wallach, J. S., Del Vecchio, M., Wilder, E. L., Zhou, H., Quinn, W. G. & Tully, T. 1994 Induction of a dominant negative CREB transgene specifically blocks long-term memory in *Drosophila*. *Cell* **79**, 49–58. (doi:10.1016/0092-8674(94)90399-9)
- 68 Yin, J. C., Wallach, J. S., Wilder, E. L., Klingensmith, J., Dang, D., Perrimon, N., Zhou, H., Tully, T. & Quinn, W. G. 1995 A *Drosophila* CREB/CREM homolog encodes multiple isoforms, including a cyclic AMP-dependent protein kinase-responsive transcriptional activator and antagonist. *Mol. Cell. Biol.* 15, 5123-5130.
- 69 Bartsch, D., Casadio, A., Karl, K. A., Serodio, P. & Kandel, E. R. 1998 CREB1 encodes a nuclear activator, a repressor, and a cytoplasmic modulator that form a regulatory unit critical for long-term facilitation. *Cell* 95, 211–223. (doi:10.1016/S0092-8674(00)81752-3)
- 70 Taubenfeld, S. M., Wiig, K. A., Monti, B., Dolan, B., Pollonini, G. & Alberini, C. M. 2001 Fornix-dependent induction of hippocampal CCAAT enhancer-binding protein b and d co-localizes with phosphorylated cAMP response element-binding protein and accompanies long-term memory consolidation. *J. Neurosci.* 21, 84–91.
- 71 Keene, A. C. & Waddell, S. 2007 Drosophila olfactory memory: single genes to complex neural circuits. Nat. Rev. Neurosci. 8, 341–345. (doi:10.1038/nrn2098)
- 72 Paul, C., Stratil, C., Hofmann, F. & Kleppisch, T. 2010 cGMP-dependent protein kinase Type I promotes CREB/CRE-mediated gene expression in neurons of the lateral amygdala. *Neurosci. Lett.* 473, 82–86. (doi:10.1016/j.neulet.2010.02.020)
- 73 Mery, F., Belay, A. T., So, A. K.-C., Sokolowski, M. B. & Kawecki, T. J. 2007 Natural polymorphism affecting learning and memory in *Drosophila*. Proc. Natl Acad. Sci. USA 104, 13 051–13 055. (doi:10. 1073/pnas.0702923104)
- 74 Reaume, C. J., Sokolowski, M. B. & Mery, F. 2010 A natural genetic polymorphism affects retroactive interference in *Drosophila melanogaster*. Proc. R. Soc. B 277, 91–98. (doi:10.1098/rspb.2010.1337)
- 75 Wang, Z., Pan, Y., Li, W., Jiang, H., Chatzimanolis, L., Chang, J., Gong, Z. & Liu, L. 2008 Visual pattern memory requires foraging function in the central complex of *Drosophila*. *Learn. Mem.* **15**, 133–142. (doi:10. 1101/lm.873008)
- 76 Tomer, R., Denes, A. S., Tessmar-Raible, K. & Arendt, D. 2010 Profiling by image registration reveals common origin of annelid mushroom bodies and vertebrate pallium. *Cell* 142, 800–809. (doi:10.1016/j.cell.2010.07.043)
- 77 Allada, R. & Sigel, J. M. 2008 Unearthing the phylogenetic roots of sleep. *Curr. Biol.* 18, R670–R679. (doi:10.1016/j.cub.2008.06.033)

- 2110 C. J. Reaume & M. B. Sokolowski Review. Gene function conservation in behaviour
- 78 Zimmerman, J. E., Naidoo, N., Raizen, D. M. & Pack, A. I. 2008 Conservation of sleep: insights from non-mammalian model systems. *Trend. Neurosci.* 31, 371–376. (doi:10.1016/j.tins.2008.05.001)
- 79 Harbison, S. T., Mackay, T. F. C. & Anholt, R. R. H. 2009 Understanding the neurogenetics of sleep: progress from *Drosophila*. *Trends Genet.* 25, 262–269. (doi:10.1016/j.tig.2009.04.003)
- 80 Crocker, A. & Sehgal, A. 2010 Genetic analysis of sleep. Genes Dev. 24, 1220–1235. (doi:10.1101/gad.1913110)
- 81 Hendricks, J. C. 2000 The need for a simple animal model to understand sleep. *Progr. Neurobiol.* 61, 339-351. (doi:10.1016/S0301-0082(99)00048-9)
- 82 Raizen, D. M., Zimmerman, J. E., Maycock, M. H., Ta, U. D., You, Y.-J., Sundaram, M. V. & Pack, A. I. 2008 Lethargus is a *Caenorhabditis elegans* sleeplike state. *Nature* 451, 569–572. (doi:10.1038/nature06535)
- 83 Tobler, I. 1983 Effect of forced locomotion on the restactivity cycle of the cockroach. *Behav. Brain Res.* 8, 351–360. (doi:10.1016/0166-4328(83)90180-8)
- 84 Sauer, S., Herrmann, E. & Kaiser, W. 2004 Sleep deprivation in honey bees. *J. Sleep Res.* 13, 145–152. (doi:10. 1111/j.1365-2869.2004.00393.x)
- 85 Hendricks, J. C., Sumei, L., Kume, K., Yin, J. C.-P., Zhaohai, Y. & Sehgal, A. 2003 Gender dimorphism in the role of cycle (BMAL1) in rest, rest regulation, and longevity in *Drosophila melanogaster*. *J. Biol. Rhythms* 18, 12–25. (doi:10.1177/0748730402239673)
- 86 Shaw, P. J., Cirelli, C., Greenspan, R. J. & Tononi, G. 2000 Correlates of sleep and waking in *Drosophila mela-nogaster*. *Science* 287, 1834–1837. (doi:10.1126/science. 287.5459.1834)
- 87 Yokogawa, T., Marin, W., Faraco, J., Pezeron, G., Applebaum, L., Zhang, J., Rosa, F., Mourrain, P. & Mignot, E. 2007 Characterization of sleep in zebrafish and insomnia in hypocretin receptor mutants. *PLoS Biol.* 5, 2379–2397. (doi:10.1371/journal.pbio. 0050277)
- 88 Hendricks, J. C., Sehgal, A. & Pack, A. I. 2001 A noncircadian role for cAMP signaling and CREB activity in *Drosophila* rest homeostasis. *Nat. Neurosci.* 4, 1108–1115. (doi:10.1038/nn743)
- 89 Graves, L. A., Hellman, K., Veasey, S., Blendy, J. A., Pack, A. & Abel, T. 2003 Genetic evidence for a role of CREB in sustained cortical arousal. *J. Neurophys.* 90, 1152–1159. (doi:10.1152/jn.00882.2002)
- 90 Langmesser, S., Franken, P., Feil, S., Emmenegger, Y., Albrecht, U. & Feil, R. 2009 cGMP-dependent protein kinase type I is implicated in the regulation of the timing and quality of sleep and wakefulness. *PLoS ONE* 4, e4238. (doi:10.1371/journal.pone.0004238)

- 91 Flames, N. & Hobert, O. 2009 Gene regulatory logic of dopamine neuron differentiation. *Nature* 458, 885–890. (doi:10.1038/nature07929)
- 92 Lichtneckert, R. & Reichert, H. 2005 Insights into the urbilaterian brain: conserved genetic patterning mechanisms in insect and vertebrate brain development. *Heredity* 94, 465–477. (doi:10.1038/sj.hdy.6800664)
- 93 Donaldson, Z. R. & Young, L. J. 2008 Oxytocin, vasopressin, and the neurogenetics of sociality. *Science* 322, 900–904. (doi:10.1126/science.1158668)
- 94 Donaldson, Z. R., Kondrashov, F. A., Putnam, A., Bai, Y., Stoinski, T. L., Hammock, E. A. & Young, J. J. 2008 Evolution of a behaviour-linked microsatellite-containing element in the 5' flanking region of the primate AVPR1A gene. *BMC Evol. Biol.* 8, 180. (doi:10.1186/ 1471-2148-8-180)
- 95 Venkatesh, B., Si-Hoe, S. L., Murphy, D. & Brenner, S. 1997 Transgenic rats reveal functional conservation of regulatory controls between the Fugu isotocin and rat oxytocin genes. *Proc. Natl Acad. Sci. USA* 94, 12462– 12466. (doi:10.1073/pnas.94.23.12462)
- 96 Weaver, I. C., Cervoni, N., Champagne, F. A., D'Alessio, A. C., Sharma, S., Seckl, J. R., Dymov, S., Szyf, M. & Meaney, M. J. 2004 Epigenetic programming by maternal behavior. *Nat. Neurosci.* 7, 847–854. (doi:10.1038/nn1276)
- 97 Weaver, I. C., Champagne, F. A., Brown, S. E., Dymov, S., Sharma, S., Meaney, M. J. & Szyf, M. 2005 Reversal of maternal programming of stress responses in adult offspring through methyl supplementation: altering epigenetic marking later in life. *J. Neurosci.* 25, 11 045– 11 054. (doi:10.1523/JNEUROSCI.3652-05.2005)
- 98 Szyf, M., Weaver, I. C., Champagne, F. A., Diorio, J. & Meaney, M. J. 2005 Maternal programming of steroid receptor expression and phenotype through DNA methylation in the rat. *Front. Neuroendocrinol.* 26, 139–162. (doi:10.1016/j.yfrne.2005.10.002)
- 99 McGowan, P. O., Sasaki, A., D'Alessio, A. C., Dymov, S., Labonté, B., Szyf, M., Turecki, G. & Meaney, M. J. 2009 Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse. *Nat. Neurosci.* 12, 342–348. (doi:10.1038/nn.2270)
- 100 Dean, A. M. & Thornton, J. W. 2007 Mechanistic approaches to the study of evolution: the functional synthesis. *Nat. Rev. Genet.* 8, 675–688. (doi:10.1038/ nrg2160)
- 101 Sommer, R. J. 2009 The future of evo-devo: model systems and evolutionary theory. *Nat. Rev. Genet.* **10**, 416–422.
- 102 Geshwind, D. H. & Konopka, G. 2009 Neuroscience in the era of functional genomics and systems biology. *Nature* 461, 908–915. (doi:10.1038/nature08537)